

EpiQuik™ Global Acetyl Histone H4K12 Quantification Kit (Fluorometric)

Cat. # P-4029

INTRODUCTION

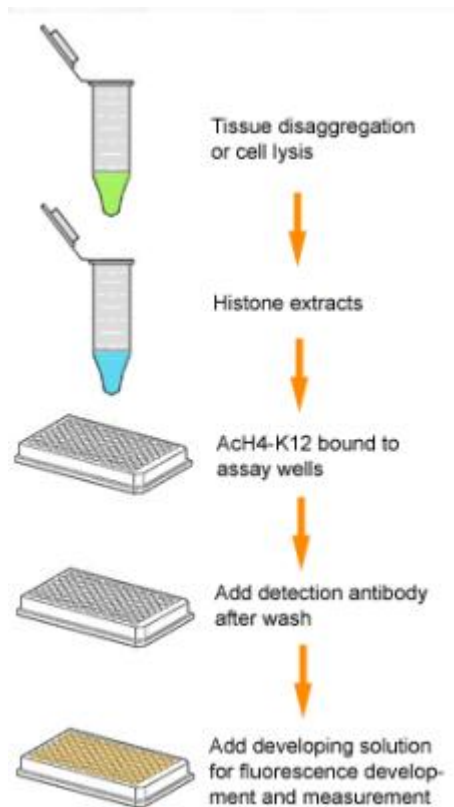
Acetylation of histones, including histone H3 and H4, have been involved in the regulation of chromatin structure and the recruitment of transcription factors to the gene promoters. Histone acetyltransferases (HATs) and histone deacetylases (HDACs) play a critical role in the control of histone H4 acetylation at multiple sites. Acetylation of histone H4 at lysine 12 (H4-K12) reflects the hyperacetylated state in histone H4 and is strongly correlated to active states of genes. H4-K12 acetylation is observed in cancer and inflammatory diseases. Histone H4-K12 acetylation may be increased by the inhibition of HDACs and decreased by HAT inhibition. Thus, quantitative detection of global acetyl histone H4-K12 would provide useful information for the better understanding of epigenetic regulation of gene activation and for developing HAT or HDAC-targeted drugs. The EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric) provides a tool for measuring global acetylation of histone H4-K12. The kit has the following features:

- Quick and efficient procedure, which can be finished within 2.5 hours.
- Innovative fluorometric assay without the need for radioactivity, electrophoresis, or chromatography.
- Specifically captures acetyl H4-K12 with the detection limit as low as 0.4 ng/well and detection range from 20 ng-5 µg/well of histone extracts.
- The control is conveniently included for the quantification of the amount of acetyl H4-K12.
- Strip microplate format makes the assay flexible: manual or high throughput.
- Simple, reliable, and consistent assay conditions.

PRINCIPLE AND PROCEDURE

The EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric) is designed for measuring global histone H4-K12 acetylation. In an assay with this kit, the acetyl histone H4 at lysine 12 is captured to the strip wells coated with an anti-acetyl H4-K12 antibody. The captured acetyl histone H4-K12 can then be detected with a labeled detection antibody, followed by a fluorescent development reagent. The ratio of acetyl H4-K12 is proportional to the intensity of fluorescence. The absolute amount of acetyl H4-K12 can be quantified by

comparing to the standard control.



Schematic Procedure for Using the EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric)

PRODUCT USE INFORMATION

The EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric) is suitable for specifically measuring global histone H4-K12 acetylation using a variety of mammalian cells (human, mouse, etc.) including fresh and frozen tissues, and cultured adherent and suspension cells.

The EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application. Suitable lab coat, disposable gloves, and eye protection are required when working with the kit.

Epigentek guarantees the performance of all products in the manner described in our product instructions.

Epigentek reserves the right to change or modify any product to enhance its performance and design.

EpiQuik™ is a trademark of Epigentek Group Inc.

The EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric) and methods of use are covered by a pending US patent.

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KIT CONTENTS

Components	48 assays P-4029-48	96 assays P-4029-96
F1 (10X wash buffer)	10 ml	20 ml
F2 (antibody buffer)	6 ml	12 ml
F3 (detecting antibody, 1 mg/ml)*	5 μ l	10 μ l
F4 (fluoro developer)*	12 μ l	24 μ l
F5 (fluoro enhancer)*	12 μ l	24 μ l
F6 (fluoro dilution)	4 ml	8 ml
Standard control (100 μ g/ml)*	10 μ l	20 μ l
Signal report solution*	5 μ l	10 μ l
Signal enhancer*	120 μ l	240 μ l
8 well sample strips (with frame)	6	9
8 well standard control strips	2	3
User guide	1	1

* For maximum recovery of the products, centrifuge the original vial prior to opening the cap.

SHIPPING AND STORAGE

Upon receipt, store F4 and standard control standard control standard control standard control at -20°C . Store all other components at 4°C away from light. The components of the kit are stable for up to 6 months from the date of shipment, when stored properly.

Note: Check if buffers F1 and F2 contain salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffers until the salts are redissolved.

MATERIALS REQUIRED BUT NOT SUPPLIED

Orbital shaker
Pipettes and pipette tips
Reagent reservoir
Fluorescence microplate reader
15 ml conical tube
1.5 ml microcentrifuge tubes

PROTOCOL

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1. a) Prepare histone extracts from cells/tissues treated or untreated by using your own successful method (acid extraction or high salt extraction).
b) For your convenience and best results, Epigentek offers the EpiQuik™ Total Histone Extraction Kit (Cat. No. OP-0006) optimized for use in the EpiQuik™ modified histone quantification series.
c) Preparation of histone extracts can also be performed using the attached procedure (See Appendix). Histone extracts can be used immediately or stored at -80°C for future use.
2. Determine the number of strip wells required. Leave these strips in the plate frame (remaining unused strips can be placed back in the bag. Seal the bag tightly and store at 4°C). Dilute F1 with distilled water (pH 7.2-7.5) at 1:9 ratio (1 ml of F1 + 9 ml of distilled water).
3. Add 50 μl of F2 into each well. For the sample, add 1-2 μg of the histone extract into the sample wells. For the standard curve, dilute standard control with F2 to 1 – 100 ng/ μl at 5-7 points (e.g., 1.5, 3, 6, 12, 25, 50, and 100 ng/ μl). Add 1 μl of standard control standard control at the different concentrations into the standard well. For the blank, do not add any nuclear extracts or standard control protein. Mix and cover the strip wells with Parafilm M and incubate at room temperature for 1 hour.

Meanwhile, prepare detection solution detection solution detection solution detection solution — for each 1 ml of detection solution to be prepared, add 1 μl of F3 and 0.5 μl of signal report solution signal report solution signal report solution signal report solution into 10 μl of diluted F1 ; mix and incubate at room temperature for 10 min. Then add 20 μl of signal enhancer signal enhancer signal enhancer signal enhancer; mix and incubate at room temperature for 15 min. Finally, add 970 μl of diluted F1 and mix.

4. Aspirate and wash the wells with 150 μl of diluted F1 3 times.
5. Add 50 μl of the detection solution detection solution detection solution detection solution to each well and incubate at room temperature for 60 min on an orbital shaker (100 rpm).
6. Aspirate and wash the wells with 150 μl of diluted F1 diluted F1 diluted F1 diluted F1 6 times.
7. Prepare fluoro development solution by adding 1 μl of F4 and 1 μl of F5 into each 400 μl of F6. Add 50 μl of fluoro - development solution into the wells and incubate at room temperature for 1-5 min away from light. The color in the standard wells containing the higher concentrations may turn slightly pink during this period. Measure and read fluorescence with a fluorescence microplate reader at 530EX/590EM nm.
Note: If the strip well frame does not fit in the fluorescence reader, transfer the solution to a standard 96-well microplate and read fluorescence at 530EX/590EM nm.

8. Calculate % histone H4-K12 acetylation:

$$\text{Acetylation \%} = \frac{\text{RFU (treated (tested) sample - blank)}}{\text{RFU (untreated (control) sample - blank)}} \times 100\%$$

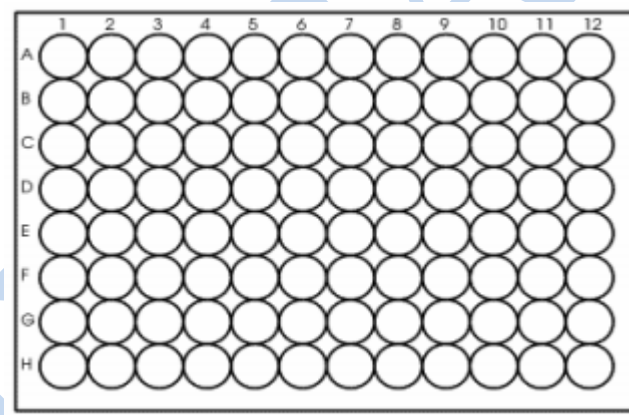
For the amount quantification, plot RFU versus amount of standard control standard control standard control and determine the slope as delta RFU/ng.

Calculate the amount of acetyl H4-K12 using the following formula:

$$\text{Amount (ng/mg protein)} = \frac{\text{RFU (sample - blank)}}{\text{Protein } (\mu\text{g})^* \times \text{slope}} \times 1000$$

* Histone extract amount added into the sample well at step 3.

Plate Configuration



- Strip 1-3 (for 96 assays) or strip 1-2 (for 48 assays) - standard wells (Labeled as SC Labeled as SC Labeled as SC Labeled as SC); the standard curve can be generated with 5-8 concentration points (includes blank).
- Example amount of standard control/well - A1: 100 ng; B1: 50 ng; C1: 25 ng; D1: 12 ng; E1: 6 ng; F1: 3 ng; G1: 1.5 ng; H1: 0 ng.
- Strip 4-12 (for 96 assays) strip 3-6 (for 48 assays) - sample wells (No label).
- Each sample or standard point can be assayed in duplicates or triplicates.

Appendix

Histone Extraction Protocol

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1. For tissues (treated and untreated), weigh the sample and cut the sample into small pieces (1-2 mm³) with a scalpel or scissors. Transfer tissue pieces to a Dounce homogenizer. Add TEB buffer (PBS containing 0.5% Triton X 100, 2 mM PMSF and 0.02% NaN₃) at 200 mg/ml, and disaggregate tissue pieces by 50-60 strokes. Transfer homogenized mixture to a 15 ml conical tube and centrifuge at 3000 rpm for 5 min at 4°C. If total mixture volume is less than 2 ml, transfer mixture to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

For cells (treated and untreated), harvest cells and pellet the cells by centrifugation at 1000 rpm for 5 min at 4°C. Resuspend cells in TEB buffer at 10⁷ cells/ml and lyse cells on ice for 10 min with gentle stirring. Centrifuge at 3000 rpm for 5 min at 4°C. If total volume is less than 2 ml, transfer cell lysates to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

2. Resuspend cell/tissue pellet in 3 volumes (approx. 200 µl/10⁷ cells or 200 mg tissues) of extraction buffer (0.5N HCl + 10% glycerol) and incubate on ice for 30 min.

3. Centrifuge at 12,000 rpm for 5 min at 4°C and remove the supernatant fraction to new vial.

4. Add 8 volumes (approx. 0.6 ml/10⁷ cells or 200 mg tissues) of acetone and leave at -20°C overnight.

5. Centrifuge at 12,000 rpm for 5 min and air-dry the pellet. Dissolve the pellet in distilled water (30-50 µl/10⁷ cells or 200 mg tissues).

6. Quantify the protein concentration. Aliquot the extract and store the extract at -20°C or -80°C.

TROUBLESHOOTING

No Signal for Both the Standard Control and the Samples

Reagents are added incorrectly.

Check if reagents are added in the proper order and if some steps of the procedure may have been omitted by mistake.

Incubation time and temperature is incorrect.

Ensure the incubation time and Temperature described in the protocol are followed correctly.

No Signal or Very Weak Signal for Only the Standard Control

The amount of standard control is not added into “standard control wells” or is added insufficiently.

Ensure sufficient amount of control is properly added to the standard control well.

No Signal for Only the Sample

The protein sample is not properly extracted.

Ensure the procedure and reagents are correct for the nuclear protein extraction.

The protein amount is added into well insufficiently.

Ensure extract contains sufficient amount of protein.

Protein extracts are incorrectly stored.

Ensure the protein extracts are stored at -20°C or -80°C .

High Background Present for the Blank

The well is not washed sufficiently.

Check if wash at each step is performed according to the protocol.

Contaminated by the standard control.

Ensure the well is not contaminated from adding the control protein or from using control protein contaminated tips.

Overdevelopment.

Decrease development time in Step 7.

ORDERING INFORMATION

Products

Size

Cat. No.

EpiQuik™ Global Acetyl Histone H4-K12 Quantification Kit (Fluorometric)

48 assays
96 assays

P-4029-48
P-4029-96

Available Related Products

Cat. No.

EpiQuik™ Global Acetyl Histone H4-K5 Quantification Kit (Colorimetric)

P-4022

EpiQuik™ Global Acetyl Histone H4-K5 Quantification Kit (Fluorometric)

P-4023

EpiQuik™ Global Acetyl Histone H4-K8 Quantification Kit (Colorimetric)

P-4024

EpiQuik™ Global Acetyl Histone H4-K8 Quantification Kit (Fluorometric)

P-4025

EpiQuik™ Global Acetyl Histone H4-K16 Quantification Kit (Colorimetric)

P-4026

EpiQuik™ Global Acetyl Histone H4-K19 Quantification Kit (Fluorometric)

P-4027

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